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Coelalysia nigriceps (Szépligeti, 1911) reared during a forensic study in Cameroon, with remarks on synonymy and biology (Hymenoptera, Braconidae, Alysiinae)

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Abstract. – *Coelalysia nigriceps* (Szépligeti, 1911) was reared as part of a study on insect scavengers in Cameroon. The nature of its possible host is discussed. *Coelalysia maculipes* (Cameron, 1912) is recognized as a new junior synonym of *C. nigriceps*. *Alysia camerunensis* Enderlein, 1912, and *Coelalysia glossinophaga* Turner, 1917, are removed from the synonymy of *C. nigriceps* and re-instated as valid species with *C. glossinophaga* as new junior synonym of *C. camerunensis* (Enderlein, 1912). Both species are illustrated, and a key is supplied to separate them. A lectotype is designated for *Coelalysia glossinophaga*.

Résumé. – ***Coelalysia nigriceps* (Szépligeti, 1911) élevé au cours d'une étude forensique au Cameroun, avec quelques observations sur sa biologie et sa synonymie (Hymenoptera, Braconidae, Alysiinae)**. *Coelalysia nigriceps* (Szépligeti, 1911) a été obtenu lors des élevages d'insectes nécrophages au Cameroun. L'identité de son hôte possible est discutée et pourrait être l'une des espèces de Calliphorides obtenues lors de l'élevage. *Coelalysia maculipes* (Cameron, 1912) est reconnu comme synonyme junior de *C. nigriceps*. La synonymie d'*Alysia camerunensis* Enderlein, 1912, et de *Coelalysia glossinophaga* Turner, 1917, avec *C. nigriceps* est rejetée et ces deux espèces sont reconnues comme valides avec *C. glossinophaga* comme nouveau synonyme junior de *C. camerunensis* (Enderlein, 1912). L'ensemble des espèces traitées est illustré et une clé d'identification est fournie. Un lectotype est désigné pour *Coelalysia glossinophaga*.

Keywords. – Alysiini, taxonomy, lectotype designation, new synonymy, rat, carrion, *Chrysomya*, Calliphoridae, *Glossina*, key.

In the hymenopteran family Braconidae Nees, 1812, Alysiinae Leach, 1815, is one of the largest cosmopolitan subfamilies (DOLPHIN & QUICKE, 2001). It contains around 2200 described valid species in 104 genera worldwide (YU *et al.*, 2009). Two tribes are usually recognized, Alysiini Leach, 1815, and Dacnusini Förster, 1862. All are koinobiont endoparasitoids of cyclorrhaphous Diptera; all Dacnusini parasitize plant-mining and gall-forming flies (WHARTON, 1997; YU *et al.*, 2009). However, Alysiini attack a wide variety of cyclorrhaphous dipteran larvae, often in decaying organic material and in moist micro-habitats.

Among the 72 valid genera currently included in the tribe Alysiini, the genus *Coelalysia* Cameron, 1911, represents a small group of nine valid species (YU *et al.*, 2009). The genus is restricted to the Afrotropical region and has been partially revised by FISCHER (1988, 2006). Host use is unknown for species of *Coelalysia* except for *C. nigriceps* (Szépligeti, 1911). It has been reared from *Musca lusoria* Wiedemann, 1824 (Diptera: Muscidae) (BRIDWELL, 1919), *Glossina sp*. (Diptera: Glossinidae) (FERRIÈRE, 1935; TURNER, 1917) and *Glossina morsitans submorsitans* Newstead, 1910 (Diptera: Glossinidae) (SIMPSON, 1918). These flies

are vectors of important human diseases, such as the human fly-related eye and diarrhoeal infections and sleeping sickness. This very broad range is from an ecological viewpoint unlikely to be correct and, therefore, the second author examined the types of *Alysia camerunensis* Enderlein, 1912, and *Coelalysia glossinophaga* Turner, 1917. For nomenclatural stability, a lectotype is designated for the last species. In addition, holotype and other specimens in the Royal Museum for Central Africa (RMCA) at Tervuren (Belgium) were examined by the first author.

MATERIALS AND METHODS

Within the framework of his PhD thesis on forensic entomology in Cameroon, the third author analysed the Diptera reared from rat carrion on the campus of the University of Yaounde I. Sixteen carcasses of rat (*Rattus norvegicus* Berkenhout, 1769, var. WISTAR) were exposed inside four wooden cages (100 ×100 × 100 cm) into the bush behind the third building of the faculty of Science at the campus (11°33'01"E - 3°51'35"N, altitude 720 m). Each cage had a 5 cm mesh for the entrance, and the cages were separated from each other by 50 m. Four cages were used as replicates and were placed under the same environmental conditions. The rats were labelled, weighed and transported to the study site. Immediately, the rats were placed inside the cages after being killed at the study site. The protocol used here for the disposition of carrion inside each cage follows CARVALHO *et al.* (2004) with some modifications. Four carcasses were exposed per cage: two were placed on top of the lattice-work deposited on a 10 cm layer of sterilized soil and two on the ground to collect the migrating and pupating maggots. The sterilized soil was put inside a transparent plastic box. Sampling of flying insects was undertaken three times daily during the first post mortem week and once a day until the remaining carcasses consisted only of bones.

After the maggots migrated into the soil beneath the carrion, the lattice-work were removed and placed in another plastic box containing 10 cm of sterilised soil. This experimental design allows collection of all the larvae migrating outside the carrion and/or pupating under it. The first box was taken back to the laboratory where it was covered by a cloth with 1 mm mesh attached with a rob around the plastic box to avoid the escape of insects. The box was placed on the shelf in the laboratory under ambient air conditions for rearing. The ambient air temperature was recorded every day with a mercury thermometer held inside the cages and in the laboratory. The boxes were checked every day, and all emerged insects were collected. For the collection the cloth was removed from the box, and the box was placed inside another one. The insects were fed with honey put on the cotton and sprayed on top of the cloth. After a feeding period of 48 hours, the insects were caught and preserved in 70% ethanol for future identification.

The identification up to the family level was done in the Zoology Laboratory of the University of Yaounde I using the identification key by DELVARE & ALBERLENC (1989). Owing to a grant offered by the Royal Museum for Central Africa, the braconid specimens could be examined by the first author using the incomplete identification keys of FISCHER (1966, 1975, 1988, 2006). Part of the specimens was sent to the second author who used unpublished information on types combined with unpublished keys to genera and species.

All the dipteran voucher specimens and part of the *Coelalysia* specimens are housed in the Zoology Laboratory of the University of Yaounde I. Other specimens of *Coelalysia* reared during our experiment are housed in the Naturalis collection (Leiden).

New records for the country are shown by *.

RESULTS

The majority of the reared insects belonged to the Diptera; three species were present in high numbers (around 70% of all specimens): *Chrysomya putoria* (Wiedemann, 1830), *Hemipyrellia fernandica* (Macquart, 1855) (both Calliphoridae) and *Hydrotae sp.* (Muscidae). The identification of the Hymenoptera revealed two species of Braconidae: 6 specimens of Microgastrinae and 100 specimens of a *Coelalysia sp.* (Alysiinae). Sixty percent of the emerged *Coelalysia* specimens were females. During some days both males and females of *Coelalysia* emerged; see records below.

With the keys by FISCHER (1988, 2006) and the redescription by PAPP (1971), females were identified as *Coelalysia nigriceps* (Szépligeti, 1911) and males as *C. maculipes* (Cameron, 1912). The identity of male specimens was confirmed by examination of the holotype of *C. maculipes* housed in RMCA. We consider the holotype of *C. maculipes* (Cameron, 1912) conspecific with *C. nigriceps* (Szépligeti, 1911) (**n. syn.**) as both were reared as opposite sexes from the same samples, and the differences concern only the colour of the metasoma and the antennae. *Coelalysia maculipes* has the fourth-seventh metasomal tergum brownish black (yellowish in *C. nigriceps*) and the antenna is entirely blackish and has no pale ring (with a whitish ring in *C. nigriceps*). The same sexual variation was found among the other specimens identified as *C. bicolor* (Szépligeti, 1911) or *C. camerunensis* (Enderlein, 1912).

The examination of the types of *Alysia camerunensis* Enderlein, 1912 (photographs of the female lectotype sent by Mr. A. Litton) and *Coelalysia glossinophaga* Turner, 1917 (syntypes on loan) revealed that they were incorrectly synonymized with *C. nigriceps* (Szépligeti, 1911). *Coelalysia camerunensis* (Enderlein, 1912) is re-instated as a valid species; we consider *C. glossinophaga* Turner, 1917, as a junior synonym (**n. syn.**) of *C. camerunensis*. The female of this last species in The Natural History Museum (London) type collection (3.c.801, Gold Coast, Northern Territories) is here designated as lectotype of *Coelalysia glossinophaga* **(present designation)** to increase nomenclatural stability.

Coelalysia nigriceps (Szépligeti, 1911) (fig. 1-12)

Idiasta nigriceps Szépligeti, 1911: 417.
Coelalysia nigriceps; PAPP, 1971: 228.
Idiasta bicolor Szépligeti, 1911: 417 (synonymized by PAPP, 1971).
Alysia maculipes Cameron, 1912: 381, **n. syn.**
Idiasta africana Szépligeti, 1914: 229 (synonymized by PAPP, 1971).

Material examined. – **Democratic Republic of Congo**. ♂ (holotype of *C. maculipes*), "Type (red label)" "Musée du Congo, Dima, 23.ix.(19)08, *A. Keller*" "R. det. W. 189" "*Alysia maculipes* Cam. Type (hand written by P. Cameron)"; 3 ♀, "Musée du Congo, Elisabethville, 5.III.1912, *Dr. Bequaert*"; ♀, *idem*, 11.II.1912; ♀, "Musée du Congo, Rutshuru, 1937, *J. Ghesquière*"; 2 ♀ "Musée du Congo, Albertville, XII.1918, *R. Mayné*"; ♂, "Musée du Congo, Béni à Lesse, fin VII.1911, *Dr. Murtula*" "*C. bicolor* Szpl. Det Dr Fahringer"; ♀, "Coll. Mus Tervuren, Mayumbe : Tshela, 13-17.II.1916, *R. Mayné*"; 2 ♂, "Musée du Congo, Mayumbe, 1917, *R. Mayné*" "*C. maculipes* Cam., det. Dr Fahringer"; 4 ♂, "Mus du Congo, Mayumba, 1917, *R. Mayné*"; ♀, "Coll. Mus Tervuren, Mayumbe, 1917, *R. Mayné*"; ♀, "Coll. Mus Tervuren, Congo belge, Mayumbe, 1917, *R. Mayné*"; ♂, "Coll. Mus Tervuren, Albertville, XII.1918, *R. Mayné*"; ♂, "Musée du Congo, Albertville, 1-20.I.1919, *R. Mayné*" "*C. maculipes* Cam., det Dr. Fahringer"; ♂, "Coll. Mus Tervuren, Bas-Uélé, VII-VIII.1920, *L. Burgeon*"; ♂, "Musée du Congo, Kasai, Makumbi, 18.X.1921, *Dr H. Schouteden*" "*C. maculipes* Cam., Dét. Dr Fahringer"; ♂, "Coll. Mus Tervuren, Congo belge, Haut-Uélé : Moto, VI.1922, *L. Burgeon*"; ♂, *idem*, IV-V.1923; ♀, *idem*, X-XI.1923; ♀, "Coll. Mus Tervuren, Liberia, Mench-Toum, 29.VIII.1926, *D. J. Bequaert*"; 6 ♂, "Coll. Mus Tervuren, Congo belge, Ituri, Sasonge ('taradje), 18.III.1930, *A. Collart*"; ♂, "Coll. Mus Tervuren, Congo belge, Kivu, Lulenga, fin IX.1932, *L. Burgeon*"; ♂, "Coll. Mus Tervuren, Congo belge, Uélé, Dingila, VII.1933, *H. J. Bredo*"; ♂, "Coll. Mus Tervuren, Congo belge, Bambesa, 30.VIII.1933, *J. V. Leroy*"; ♀, *idem*, 15.IX.1933; ♀, *idem*, 20.X.1933; ♀, *idem*, 30.X.1933; ♂, "Coll. Mus Tervuren, Congo belge, Kivu, Rutshuru (riv. Musugereza)

1100 m, 4.VII.1935, *G. F. De Witte*, 1608"; ♀, "Coll. Mus Tervuren, Kivu: Rutshuru riv. Kanzawe, 1200 m, 16.VII.1935, *G. F. de Witte*: 1658"; ♂, "Coll. Mus Tervuren, Congo belge, Rutshuru, 20.II.1936, *L. Lippens*"; ♂, "Coll. Mus Tervuren, Congo belge, Kivu, Sake, 14.III.1936, *L. Lippens*"; 2 ♂, "Coll. Mus Tervuren, Ruanda, L. Nyakibugu, 17.III.1936, *L. Lippens*"; 2 ♂, "Coll. Mus Tervuren, Congo belge, Rutshuru, 12.V.1936, *L. Lippens* (150)"; 2 ♂, "Coll. Mus Tervuren, Congo belge, Rutshuru, XII.1937, *J. Ghesquière*"; ♂, "Coll. Mus Tervuren, Congo belge, Mongbwalu (Kibo), 1938, *Mme Scheitz*"; ♂, "Coll. Mus Tervuren, Congo belge, PNG, Miss. H. De Saeger, PpK 73/d/9, 8.IV.(19)52, *H. De Saeger* 3311"; ♀, "Coll. Mus Tervuren, Terr. De Dibaya, Kamponde, 1945, *Rev. Fr. Allaer*"; ♀, "Coll. Mus Tervuren, Haut Uélé, Paulis, XII.1947, *P. L. G. Benoit*"; ♀, "Coll. Mus Tervuren, Elisabethville (à la lumière), II.1949, *Ch. Seydel*"; ♀, *idem*, V.1949; ♀, *idem*, XI.1949; ♀, *idem*, III.1950; ♀, *idem*, IX.1950; ♀, *idem*, IX.1959; ♀, *idem*, I.1960; ♂, "Coll. Mus Tervuren, Congo belge, Costermansville, IV-V.1949, *H. Bomans*"; ♂ and ♀, *idem*, VIII.1949; ♂, *idem*, 1951; 2 ♂, "Coll. Mus Tervuren, Congo belge, Kivu, Ibonda, 1952, *M. Vandelannoite*"; ♀, "Coll. Mus Tervuren, Congo belge, Equateur: Bokuma, VII.1952, *R. P. Lootens*"; ♂, "Coll. Mus Tervuren, PNG, Miss H. De Saeger, Mabanga, 29.IX.1952, *H. De Saeger* 4103"; ♂, "Coll. Mus Tervuren, Tshuaba: Bamania, 9.X.1952, *R. P. Hulstaert*"; ♀, "Coll. Mus Tervuren, Tshuapa, Eala, 8.XII.1952, *P. Basilewsky*"; ♂, "Coll. Mus Tervuren, Ruanda, Rukoma (Cheff.) terr; Nyanza, I.1953, *J. P. Basilewsky*"; ♂, *idem*, II. 1953; 4 ♂, "Coll. Mus Tervuren, Urundi: Buruni, 1800-2000 m, 5-12.III.1953, *J. P. Basilewsky*"; ♂, "Coll. Mus Tervuren, Congo belge, PNA, 12.III.1953, *P. Vanschuytbroek & H. Synove* 7703-04, Secteur Tshiaberimu, Fl. Talia Nord, 2340m"; ♂, "Coll. Mus Tervuren, Congo belge, PNA, 26-28.VIII.1953, *P. Vanschuytbroek & V. Hendricks* 4999-5005, Secteur Tshiaberimu, riv. Mbulikerere affl. Talia Nord, 2720 m"; ♂, "Coll. Mus Tervuren, Congo belge, PNA, 27.VIII.1953, *P. Vanschuytbroek & V. Hendricks* 5009-11, Secteur Tshiaberimu, riv. Kalivina affl. Talia Nord, 2720 m"; ♂, "Coll. Mus Tervuren, Congo belge, PNA, 23.III.1954, *P. Vanschuytbroek & H. Synove* 7705-12, Secteur Tshiaberimu, Fl. Talia Nord, 2340 m"; ♂, "Coll. Mus. Congo, Tshuaba, Ikela, 1955, *R. P. Lootens*"; 1 ex., "Coll. Mus Tervuren, Congo belge, PNA, 12-17.VII.1955, *P. Vanschuytbroek*, 13741-47, Mont Hoyo, grotte Yolokafiri, 1030 m"; ♀, "Coll. Mus Tervuren, Congo Belge, P.N.A., 9.XI.1956, *P. Vanschuytbroeck* VS798" "Massif Ruwenzori, tête de source riv. Indray affl. Semliki, 1840 m"; ♂, "Coll. Mus Tervuren, Congo belge, Lualaba, Ruwe (piège lumineux), II.1960, *Dr. V. Allard*"; ♀, "Coll. Mus Tervuren, Lualaba, Ruwa (piège lumineux), II.1960, *Dr. V. Allard*"; ♂, "Coll. Mus Tervuren, Congo belge, Uélé, *De Greef*"; ♂, "Coll.

Fig. 1. – *Coelalysia nigriceps* (Szépligeti), ♀, Cameroon, Yaoundé. Habitus, lateral aspect.

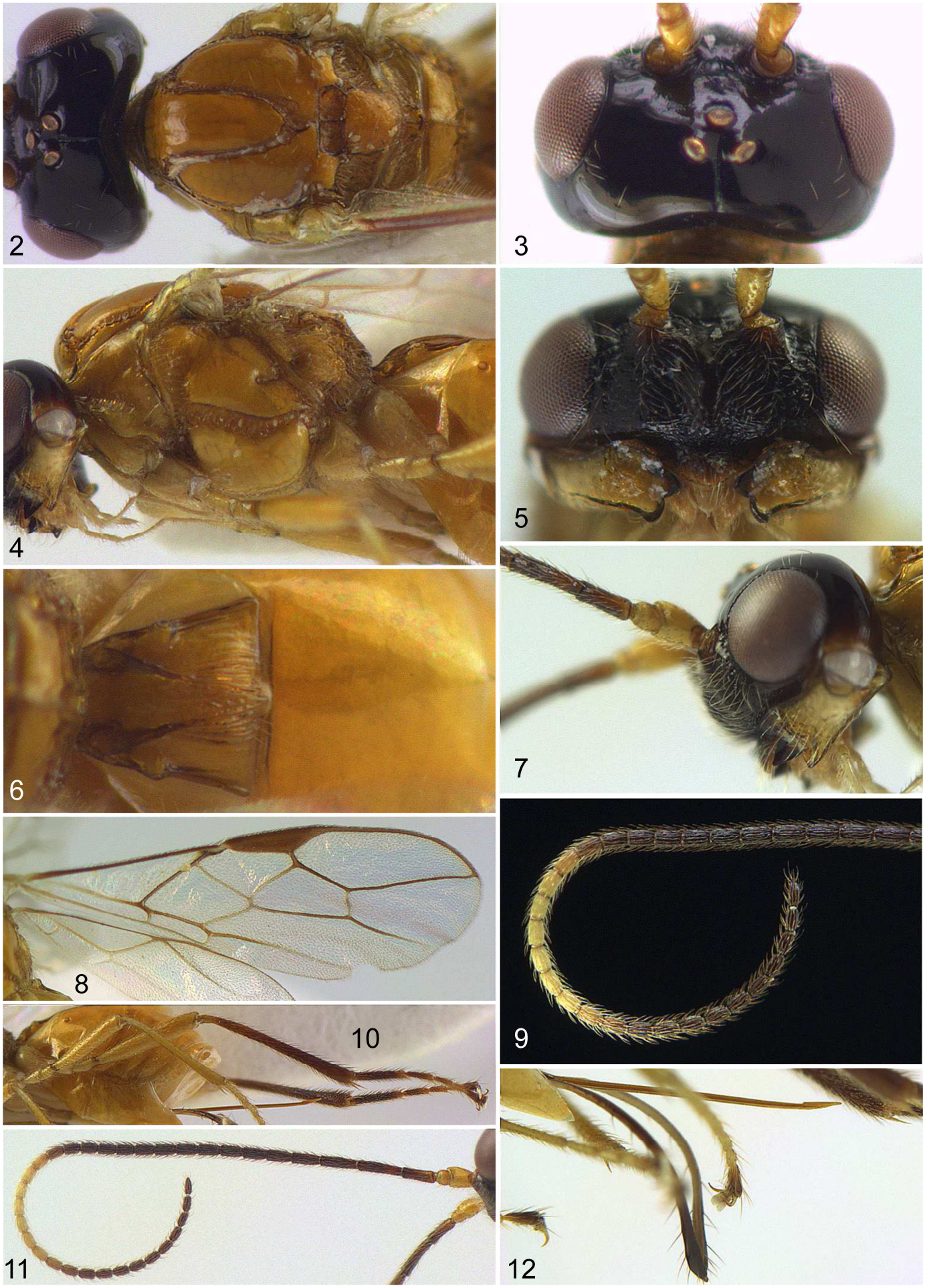

Fig. 2-12. – *Coelalysia nigriceps* (Szépligeti), ♀, Cameroon, Yaoundé. – 2, Head and mesosoma, dorsal aspect. – 3, Head, dorsal aspect. – 4, Mesosoma, lateral aspect. – 5, Head, anterior aspect. – 6, First and second metasomal tergites, dorsal aspect. – 7, Head, lateral aspect. – 8, Wings. – 9, Apical half of antenna. – 10, Hind leg. – 11, Antenna. – 12, Ovipositor and its sheath.

Mus Tervuren, Congo belge, Uélé, *Degreef*". **Cameroon**. 3 ♂, "Coll. Mus Tervuren, Cameroun: Nkobilsson, dept. Nyong-Sanaga, X.(19)63, *L. G. Segers leg.*". Data collected during our study [the abbreviation "ex" is used for "emergence date"]: "Cameroun, Yaoundé, Campus UYI, *Col. Feugang Youmessi*": 2 ♀, 29.XI.2010, cage A; 2 ♀, 30.XI.2010, cage A; 11 ♀, 30.XI.2010, cage A; 2 ♂, 1.XII.2010, cage A; 10 ♀, 1.XII.2010, cage B; 2 ♀, 2.XII.2010, cage B; ♀, 4.XII.2010, ex: 6.I.2011, cage A; ♂, 4.XII.2010, ex: 18.XII.2010, cage A; 12 ♂, 4.XII.2010, ex: 29.XII.2010, cage A; 2 ♀ and 1 ♂, 4.XII.2010, ex: 30.XII.2010, cage A; 9 ♀, 15.XII.2010, cage C; 4 ♀, 15.XII.2010, cage D; ♀, 18.XII.2010, ex: 29.XII.2010, cage D; 1 ♀ and 13 ♂, 18.XII.2010, ex: 25.XII.2010, cage B; 4 ♀, 18.XII.2010, ex: 27.XII.2010, cage B; 2 ♂, 18.XII.2010, ex: 6.I.2011, cage D; 1 ♀ and 2 ♂, 18.XII.2010, ex: 8.I.2011, cage D; 1 ♀ and 2 ♂, 18.XII.2010, ex. 10.I.2011, cage C; 2 ♀ and 1 ♂, 18.XII.2010, ex: 12.I.2011, cage C; 4 ♀ and 2 ♂, 18.XII.2010, ex: 14.I.2011, cage C; 2 ♀ and 1 ♂, 18.XII.2010, ex : 15.I.2011, cage C; 3 ♀, 18.XII.2010, ex: 17.I.2011, cage C; 4 ♂, 18.XII.2010, ex: 27.XII.2010, cage B. **Burundi**. ♂, "Coll. Mus Tervuren, Urundi, Bururi, 1800-2000m, 5/12.III.1953, *P. Basilewsky*". **Ivory Coast**. ♀, "Coll. Mus Tervuren, Côte d'Ivoire: Bingerville, 1/15.IX.1963, *J. Decelle*"; ♂, *idem*, VI.1962; ♂, *idem*, X.1962; ♀, *idem*, XI.1962. **Nigeria**. ♀, "Coll. Mus Tervuren, Nigeria: Bida CRH, N-W State, 1.IX.1970, *col. J. T. Medier*".

Distribution. – Burundi*, Cameroon, Democratic Republic of Congo, Equatorial Guinea, Guinea, Ivory Coast*, Kenya, Malawi, Mozambique, Nigeria*, Tanzania, Togo, Uganda.

Coelalysia camerunensis (Enderlein, 1912), **stat. rev.** (fig. 13-25)

Alysia camerunensis Enderlein, 1912: 99.
Coelalysia camerunensis; Turner, 1917: 177.
Coelalysia glossinophaga Turner, 1917: 177, **n. syn.**

As already suspected because of the different host use (*Coelalysia glossinophaga* was reared as a parasitoid of *Glossina spp.*), this species is not conspecific with *C. nigriceps*. The female lectotype of *C. glossinophaga* has 15th-28th antennal flagellomeres white (of total of 40); the clypeus is subtriangular, the notauli are finely crenulate and only the head is black, remainder yellowish brown (fig. 13), length of the body 5 mm, of the ovipositor sheath 2 mm and the first tergite is as long as wide. The antenna of the male paralectotype of *C. glossinophaga* is dark brown. According to the redescription of *C. nigriceps* by PAPP (1971), the clypeus is triangular and rather acute. According to FISCHER (1988), *C. nigriceps* has the clypeus rounded ventrally; probably he mixed the species up with *C. camerunensis*. Both species can be separated as follows.

Fig. 13. – *Coelalysia camerunensis* (Enderlein), ♀ lectotype of *C. glossinophaga* Turner, Ghana. Habitus, lateral aspect.

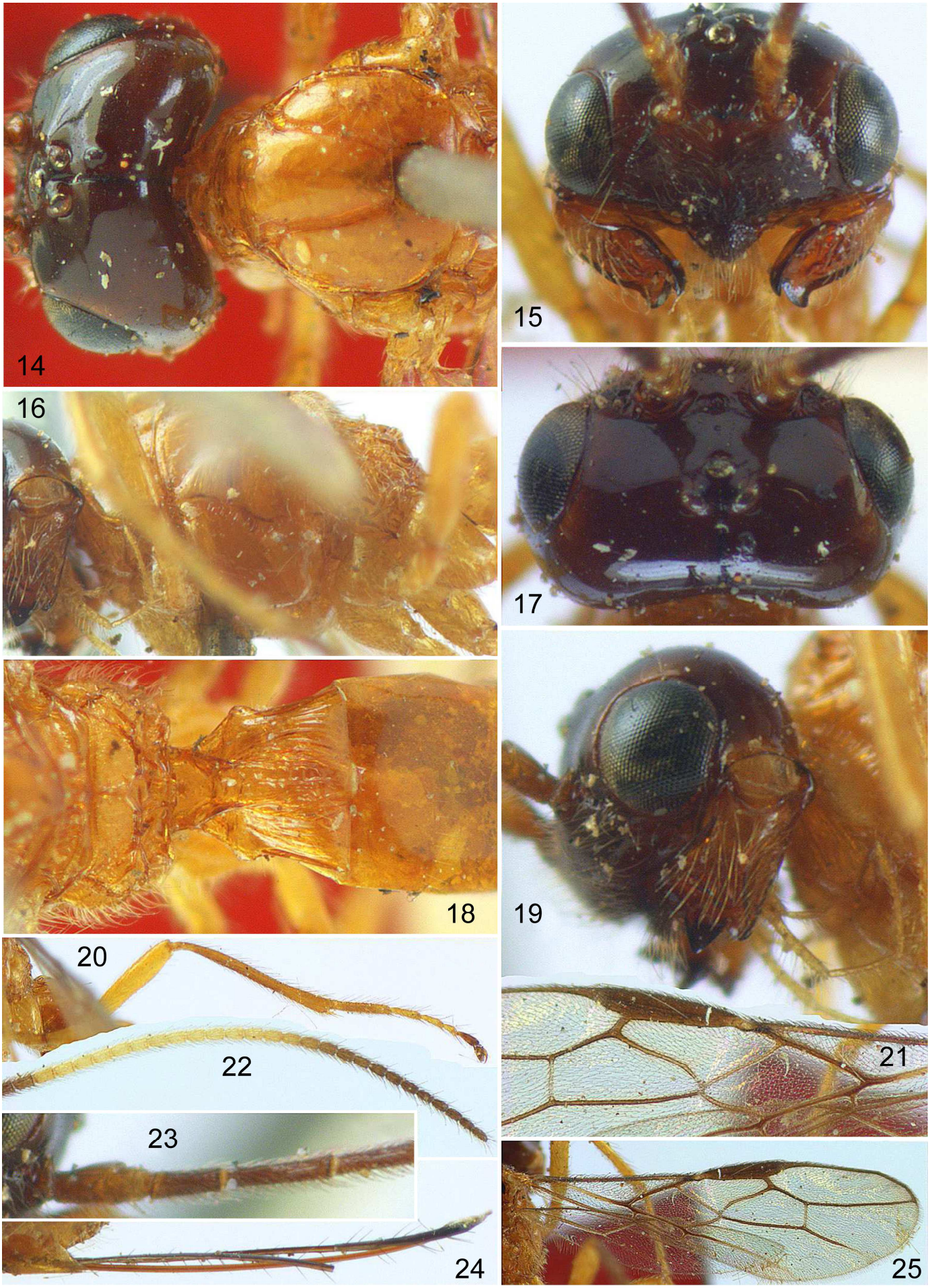

Fig. 14-25. – *Coelalysia camerunensis* (Enderlein), ♀ lectotype of *C. glossinophaga* Turner, Ghana. – 14, Head and mesosoma, dorsal aspect. – 15, Head, anterior aspect. – 16, Mesosoma, lateral aspect. – 17, Head, dorsal aspect. – 18, Propodeum, first and second metasomal tergites, dorsal aspect. – 19, Head, lateral aspect. – 20, Hind leg. – 21, Detail of fore wing. – 22, Antenna. – 23, Basal segments of antenna. – 24, Ovipositor and its sheath. – 25, Wings.

– Vein 2-SR of fore wing 0.9-1.0 times as long as vein 3-SR (fig. 8); antenna of female with 5-8 ivory or white flagellomeres (and with 0-2 brown segments; fig. 9, 11); clypeus more or less rounded ventrally (fig. 5); vein 3-CU1 of fore wing slender and comparatively long (fig. 8); precoxal sulcus distinctly widened medially (fig. 4); area in front of anterior ocellus with small isolated pit (fig. 3, 5); middle tooth of mandible comparatively slender, basally narrower than its lateral sides (fig. 7); setose part of ovipositor sheath 0.4-0.5 times as long as fore wing, 0.8-1.0 times as long as hind tibia and 0.8-0.9 times as long as metasoma (fig. 1, 12); parasitoid of necrophilous dipterous larvae in carrion; East and West Africa ..... ***C. nigriceps*** (Szépligeti)
– Vein 2-SR of fore wing 1.5-1.6 times as long as vein 3-SR (fig. 21); antenna of female with about 14 ivory or white flagellomeres (fig. 22); clypeus distinctly acute ventrally and triangular (fig. 15); vein 3-CU1 of fore wing widened and comparatively short (fig. 21); precoxal sulcus comparatively narrow medially (fig. 16); area in front of anterior ocellus with only a shallow depression connected to ocellus (fig. 17); middle tooth of mandible comparatively robust, with about equal sides (fig. 19); setose part of ovipositor sheath 0.5-0.6 times as long as fore wing, 1.4-1.5 times as long as hind tibia and 1.1-1.2 times as long as metasoma (fig. 13, 24); parasitoids of *Glossina spp.*; West Africa: Cameroon, Ghana ..... ***C. camerunensis*** (Enderlein)

CONCLUSION

Our study shows that *Coelalysia nigriceps* is rather parasitoid of one dipterous larvae developing on rat carrion and not a parasitoid of *Glossina* species. Only individual rearing of necrophagous dipteran larvae could formally determine the exact host. But considering the size of the adult parasitoid and of the emerged flies, the Calliphoridae species [such as *Chrysomya putoria* (Wiedemann, 1830) or *Hemipyrellia fernandica* (Macquart, 1855)] should be some good candidates as potential host. Among the females collected around the carrion, only the sarcophagids (but no larva were reared during this experiment) could be also considered as potential host. The other species of reared diptera have smaller size that could not be enough to allow a full development of the *Coelalysia* species. The known distribution of *C. nigriceps* is extended with Burundi, Ivory Coast and Nigeria. The parasitoid of *Glossina spp.* belongs to another species, *C. camerunensis*, and is only known from West Africa.

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Karoline FRITZSCHE et Alex DELOBEL. – ***Megabruchidius dorsalis* (Fåhraeus, 1839), Bruche nouvelle pour la faune française (Col., Chrysomelidae, Bruchinae)**

En 2008, DELOBEL & DELOBEL signalent la présence en France métropolitaine (Montpellier) d'une Bruche d'origine asiatique, *Megabruchidius tonkineus* (Pic, 1904), dans des graines de févier d'Amérique, *Gleditsia triacanthos* L. Nous avons obtenu une autre espèce du genre *Megabruchidius* Borowiec, 1984, *M. dorsalis*, de graines de févier d'Amérique récoltées le 21 mai 2011 à Paris, sur l'avenue des Champs-Elysées, puis le 19 novembre 2011 dans le parc départemental de Sceaux (Hauts-de-Seine). Cette espèce, répandue de l'Inde au Japon et jusqu'en Papouasie-Nouvelle-Guinée, a été signalée à deux reprises en Europe : en Italie (MIGLIACIO & ZAMPETTI, 1989), puis en Hongrie et en Suisse (YUS RAMOS, 2009). Sa présence en France, dans la région parisienne, ne doit pas surprendre, dans la mesure où l'une de ses plantes hôtes, *G. triacanthos,* est abondamment plantée dans nos rues et nos parcs.

Après environ un mois d'incubation au laboratoire, à température ambiante, environ 1000 bruches, dont seulement deux mâles et trois femelles de *M. dorsalis,* ont émergé de la cinquantaine de gousses récoltées à Paris en mai. Tous les autres spécimens appartenaient à l'espèce *M. tonkineus*. Comme à Budapest (YUS RAMOS, 2009), les deux espèces coexistent donc à Paris sur les mêmes hôtes. Des douze gousses récoltées à Sceaux en novembre, trois mâles et une femelle de *M. dorsalis* ont émergé en décembre et janvier 2012. Aucun individu de *M. tonkineus* n'a été obtenu de ces gousses. La durée du développement larvaire est probablement très variable et largement dépendante des températures ambiantes. Il semble que plusieurs générations puissent se succéder (en se chevauchant) dans les gousses qui restent disponibles dans les arbres ou au sol une grande partie de l'année, au moins de novembre à mai ou juin.